

---

 Communications to the Editor
 

---

 ISOLATION AND STRUCTURAL  
 ELUCIDATION OF STIPIAMIDE,  
 A NEW ANTIBIOTIC EFFECTIVE  
 TO MULTIDRUG-RESISTANT  
 CANCER CELLS

Sir:

Resistant of tumor cells to multiple cytotoxic drugs is a major impediment to cancer chemotherapy. Multidrug-resistance (MDR) is characterized by decreased cellular sensitivity to anticancer agents due to the increased expression of a membrane glycoprotein with a MW of approximately 170 kd, termed P-glycoprotein<sup>1</sup>.

It is well known that colchicine-resistance cells have simultaneously acquired resistance to doxorubicin and vinblastine<sup>2</sup>. Thus, it could be expected that compounds active against colchicine-resistance cells will be useful for overcoming MDR to antitumor substances.

By employing colchicine-resistance KB(CH<sup>R</sup>)<sup>3</sup> cells, we undertook the screening for agents which were more active to KB(CH<sup>R</sup>) cells in the presence of colchicine<sup>†</sup>. As a result, a gliding bacterium isolated from a soil sample from Izu Peninsula, Japan was found to produce a new antibiotic named stipiamide. The producing organism was identified as *Myxococcus stipitatus* (AJ-12587) by taxonomic comparison with type strains of *Myxococcus* sp. as well as by its morphological, cultural and physiological characteristics. In this paper, we will report the fermentation, isolation and structure elucidation of stipiamide.

The organism was cultivated in 5-liter Sakaguchi flasks containing 1 liter of the Casitone liquid medium consisting of Casitone (Difco) 2% and MgSO<sub>4</sub> 0.2% (pH 7.2). The flasks were incubated at 28°C for 4 days on a reciprocal shaker at 115 rpm. In this medium, the organism grew as a homogeneous cell suspension. The cells harvested by centrifugation from the whole broth (5 liters) were stirred with 500 ml of acetone. The solvent extract was concentrated *in vacuo* to a small volume and the active material was extracted from the aqueous

residue twice with each 100 ml of EtOAc. The separated organic layer was dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo* to give a dark brownish oil, which was subjected to silica gel column chromatography (Wakogel C-200, 5 × 10 cm) packed in CHCl<sub>3</sub>. After developing with 800 ml of CHCl<sub>3</sub>, the column was developed by a mixture of CHCl<sub>3</sub>-MeOH (49:1). The active fractions were combined and concentrated *in vacuo* to afford a crude oil. Further purification of the oil was achieved by preparative TLC (Merck, Kieselgel 60F<sub>254</sub> Art. No. 5744, 0.5 mm) developed with CHCl<sub>3</sub>-MeOH (21:1). The active band was collected and purified once more by preparative TLC developed with hexane-EtOAc (2:3) to yield 106 mg of stipiamide.

Its physico-chemical properties are shown in Table 1. The molecular formula of stipiamide was determined to be C<sub>32</sub>H<sub>45</sub>NO<sub>3</sub> by HRFAB-MS data (*m/z* 492.3416 (M+H)<sup>+</sup>, Calcd: 492.3477).

The <sup>13</sup>C and <sup>1</sup>H NMR spectral data of stipiamide are summarized in Table 2. The functionalities of the carbons in stipiamide were revealed by DEPT experiments to be as follows; CH<sub>3</sub> × 6, CH<sub>2</sub> × 2, CH × 16, CH<sub>2</sub>O × 1, CHO × 1, CHN × 1, C=O × 1 and C × 4.

The <sup>13</sup>C resonance at δ<sub>c</sub> 169.8 was assigned to an amide carbonyl due to an IR absorption band at 1645 cm<sup>-1</sup>. The partial structures shown in Fig. 1 were determined by analysis of the DQF-COSY spectrum which showed the sequences of 1'-H to 2'-H (I), 3-H to 9-H (II), 11-H to 13-H (III), 15-H

Table 1. Physico-chemical properties of stipiamide.

Appearance	Yellowish oil
Molecular formula	C <sub>32</sub> H <sub>45</sub> NO <sub>3</sub>
HRFAB-MS ( <i>m/z</i> )	492.3416 (M+H) <sup>+</sup> , Calcd: 492.3477
UV λ <sub>max</sub> <sup>MeOH</sup> nm (ε)	207 (9,820), 264 (6,730), 359 (31,500)
λ <sub>max</sub> <sup>MeOH-NaOH</sup> nm (ε)	209 (32,200), 264 (6,780), 359 (31,300)
IR ν <sub>max</sub> <sup>CCl4</sup> (cm <sup>-1</sup> )	3375, 2950, 1645, 1505, 1455, 995, 965, 700
[α] <sub>D</sub> <sup>23</sup>	-115.1° (c 1.45, MeOH)
Solubility	Soluble in CCl <sub>4</sub> , CHCl <sub>3</sub> , MeOH Insoluble in H <sub>2</sub> O
Rf (Kieselgel 60 F <sub>254</sub> )	0.28 (CHCl <sub>3</sub> -MeOH, 21:1) 0.37 (Hexane-EtOAc, 2:3)

<sup>†</sup> IC<sub>50</sub> of colchicine to KB(CH<sup>R</sup>) cells was 8.5 μg/ml, while that to normal KB cell was 0.08 μg/ml. KB(CH<sup>R</sup>) cells were about 30 times as resistant to daunorubicin as KB cell.

Table 2.  $^{13}\text{C}$  and  $^1\text{H}$  NMR signals of stipiamide taken in acetone- $d_6$ .

No.	$\delta_{\text{C}}$	$\delta_{\text{H}}$
1	169.8	
2	132.2	
3	133.8	7.00 (1H, d, $J=11$ Hz)
4	129.4	6.58 (1H, dd, $J=11, 15$ Hz)
5	134.2	7.08 (1H, dd, $J=10, 15$ Hz)
6	129.3	6.10 (1H, dd, $J=10, 10$ Hz)
7	133.5	6.15 (1H, dd, $J=10, 10$ Hz)
8	122.9	6.78 (1H, dd, $J=10, 15$ Hz)
9	141.8	6.39 (1H, d, $J=15$ Hz)
10	135.2	
11	140.0	5.64 (1H, d, $J=10$ Hz)
12	38.1	2.80 (1H, qdd, $J=7, 7, 10$ Hz)
13	82.2	3.84 (1H, dd, $J=5, 7$ Hz)
14	137.0	
15	133.5	5.25 (1H, d, $J=10$ Hz)
16	32.8	2.43 (1H, m)
17	40.8	1.50, 1.63 (2H, m)
18	34.9	2.50, 2.60 (2H, m)
19	144.0	
20	129.6	7.16 (1H, d, $J=8$ Hz)
21	129.4	7.23 (1H, t, $J=8$ Hz)
22	126.7	7.13 (1H, t, $J=8$ Hz)
23	129.4	7.23 (1H, t, $J=8$ Hz)
24	129.6	7.16 (1H, d, $J=8$ Hz)
2-CH <sub>3</sub>	13.7	1.96 (3H, d, $J=8$ Hz)
10-CH <sub>3</sub>	13.5	1.87 (3H, s)
12-CH <sub>3</sub>	18.7	0.95 (3H, d, $J=7$ Hz)
14-CH <sub>3</sub>	13.0	1.60 (3H, s)
16-CH <sub>3</sub>	22.0	0.96 (3H, d, $J=7$ Hz)
1'	49.0	4.05 (1H, qt, $J=7, 7$ Hz)
2'	66.9	3.54 (2H, m)
1'-CH <sub>3</sub>	17.8	1.17 (3H, s)

to 18-H (IV), and a phenyl ring (V).

The complete connectivity of the carbon skeleton of the polyene chain and phenethyl residue could easily be established by an HMBC experiment.<sup>4,5)</sup> Thus, the  $^{13}\text{C}$ - $^1\text{H}$  long range couplings observed with methyl groups in Fig. 2 (2-CH<sub>3</sub> ( $\delta_{\text{H}}$  1.96), 10-CH<sub>3</sub> ( $\delta_{\text{H}}$  1.87) and 14-CH<sub>3</sub> ( $\delta_{\text{H}}$  1.60)) enabled the partial structures (I) to (IV) in Fig. 1 separated by quaternary carbons to be connected to form a highly unsaturated chain. In addition, the couplings between aromatic protons (20-H, 24-H ( $\delta_{\text{H}}$  7.16)) and benzyl methylene (18-H ( $\delta_{\text{H}}$  2.50, 2.60)) proved the partial structure (IV) shown in Fig. 2. Based on these results, the total planar structure of stipiamide can be depicted as shown in Fig. 3.

Stereochemical studies were facilitated by analysis of the  $^1\text{H}$ - $^1\text{H}$  coupling constants (3-H to 9-H) (Fig. 1 (II)) and one dimensional NOE difference experiments. NOEs obtained by irradiating at methyl protons (2-CH<sub>3</sub> ( $\delta_{\text{H}}$  1.96), 10-CH<sub>3</sub> ( $\delta_{\text{H}}$  1.87) and 14-CH<sub>3</sub> ( $\delta_{\text{H}}$  1.60)) were as follows; (2-CH<sub>3</sub>→4-H ( $\delta_{\text{H}}$  6.58), N-H ( $\delta_{\text{H}}$  6.89); 10-CH<sub>3</sub>→8-H ( $\delta_{\text{H}}$  6.78), 12-H ( $\delta_{\text{H}}$  2.80), and 14-CH<sub>3</sub>→12-H ( $\delta_{\text{H}}$  2.80), 16-H ( $\delta_{\text{H}}$  2.43)). These results established the stereochemistries of the double bonds in stipiamide as (2*E*,4*E*,6*Z*,8*E*,10*E*,14*E*). This structure was supported by the upfield  $^{13}\text{C}$  chemical shifts of the three allylic methyl carbons (2-CH<sub>3</sub> ( $\delta_{\text{C}}$  13.7), 10-CH<sub>3</sub> ( $\delta_{\text{C}}$  13.5), 14-CH<sub>3</sub> ( $\delta_{\text{C}}$  13.0)) ( $\gamma$ -effect<sup>6)</sup>.

The structure of stipiamide thus established is similar to that of myxalamide B<sup>7)</sup>, an alaninol amide with a highly unsaturated branched fatty acid. The chemical shifts of C-1' to C-2' and C-1 to C-15 agreed very well with those of the corresponding carbons

Fig. 1. Partial structures of stipiamide as revealed by DQF-COSY.

Values show  $^1\text{H}$ -chemical shifts in ppm. Curves indicate coupling constants in Hz.

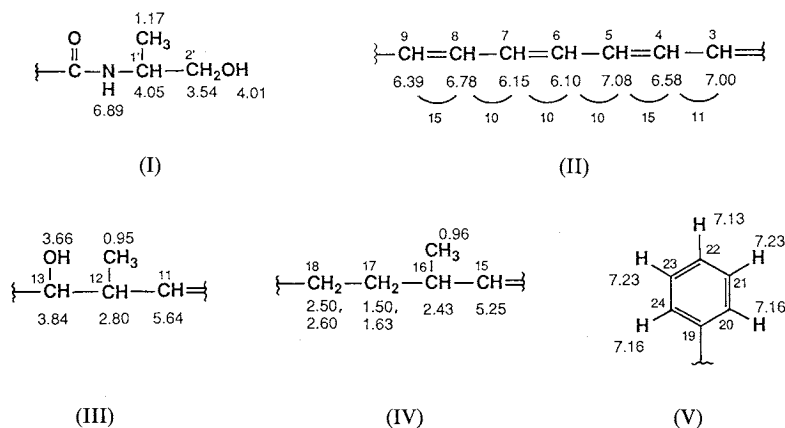


Fig. 2. Partial structures of stipiamide as revealed by HMBC.

Values and those in parentheses show  $^{13}\text{C}$ - and  $^1\text{H}$ -chemical shifts in ppm, respectively. Arrows mean long range  $^{13}\text{C}$ - $^1\text{H}$  couplings.

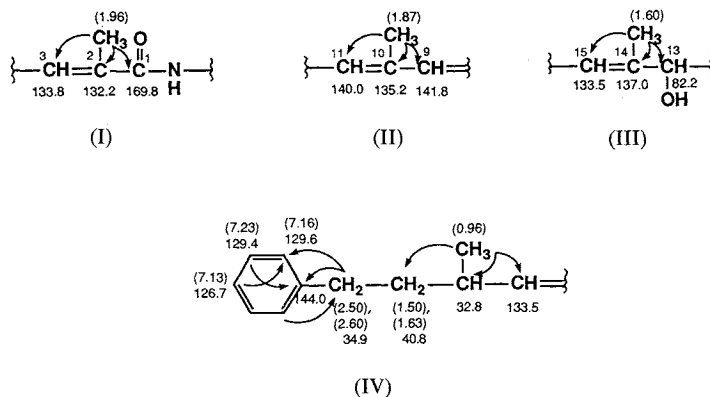
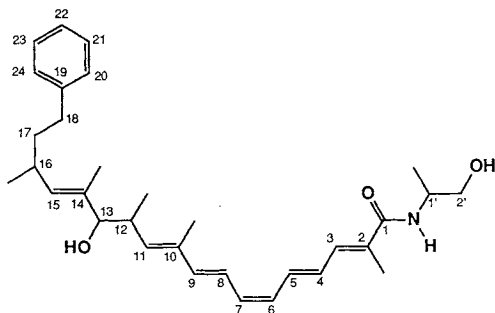


Fig. 3. Structure of stipiamide.



in myxalamide B. They differ from each other in that the terminal methyl group of the long chain in myxalamide B is replaced by a phenethyl group in stipiamide.

Stipiamide had no inhibitory effect on bacteria or yeast at a concentration of 1,000  $\mu\text{g}/\text{ml}$ , while it showed weak activity against *Pyricularia oryzae* at the same concentration. Stipiamide inhibited the growth of KB cells and its colchicine-resistant mutant KB(CH<sup>R</sup>) dose dependently in a colchicine free medium with IC<sub>50</sub> being 1.51  $\mu\text{g}/\text{ml}$  and 1.40  $\mu\text{g}/\text{ml}$ , respectively. On the other hand, stipiamide showed stronger cytotoxic activity against KB(CH<sup>R</sup>) at IC<sub>50</sub> of 0.35  $\mu\text{g}/\text{ml}$  in a medium containing 1.5  $\mu\text{g}/\text{ml}$  of colchicine. However, it showed slight effect to reduce the IC<sub>50</sub> value against the same cell (from 1.40 to 0.8  $\mu\text{g}/\text{ml}$ ) in the presence of 5  $\mu\text{g}/\text{ml}$  of doxorubicin. These experimental results may suggest that stipiamide possesses a strong synergistic effect to colchicine against KB(CH<sup>R</sup>) cell but does not affect MDR. Further

studies will be required to reveal the action mechanism of stipiamide.

#### Acknowledgments

We wish to express our hearty thanks to Prof. S. AKIYAMA of Kagoshima Univ. for providing us with KB(CH<sup>R</sup>) cells. This work was supported in part by a Grant-in-Aid for Cancer Research, The Ministry of Education, Science and Culture, Japan.

YOON JEONG KIM  
KAZUO FURIHATA<sup>†</sup>  
SHIGERU YAMANAKA<sup>††</sup>  
RYOSUKE FUDO<sup>††</sup>  
HARUO SETO\*

Institute of Applied Microbiology,  
The University of Tokyo,  
Bunkyo-ku, Tokyo 113, Japan

<sup>†</sup> Department of Agricultural Chemistry,  
Faculty of Agriculture,  
The University of Tokyo,  
Bunkyo-ku, Tokyo 113, Japan

<sup>††</sup> Central Research Laboratories,  
Ajinomoto Co., Ltd.,  
Suzukicho, Kawasaki-ku, Kawasaki 210,  
Japan

(Received January 16, 1991)

#### References

- BRADLEY, G.; P. F. JURANKA & V. LING: Mechanism of multidrug resistance. *Biochim. Biophys. Acta* 948: 87~128, 1988
- SHEN, D.; C. CARDARELLI, J. HWANG, M. CORNWELL,

- N. RICHERT, S. ISHII, I. PASTAN & M. M. GOTTESMAN: Multiple drug-resistant human KB carcinoma cells independently selected for high level resistance to colchicine, adriamycin, or vinblastine show changes in expression of specific proteins. *J. Biol. Chem.* 261: 7762~7770, 1986
- 3) SHIRAIISHI, N.; S. AKIYAMA, M. NAKAGAWA, M. KOBAYASHI & M. KUWANO: Effect of bisbenzylisoquinoline (biscoclaurine) alkaloids on multidrug resistance in KB human cancer cells. *Cancer Res.* 47: 2413~2416, 1987
- 4) SETO, H.; K. FURIHATA & M. OHUCHI: Particular utility of the HMBC technique to polypropionate derived metabolites as exemplified by erythromycin. *A. J. Antibiotics* 41: 1158~1160, 1988
- 5) BAX, A. & M. F. SUMMERS:  $^1\text{H}$  and  $^{13}\text{C}$  assignments from sensitivity-enhanced detection of heteronuclear multiple-bond connectivity by 2D multiple quantum NMR. *J. Am. Chem. Soc.* 108: 2093~2094, 1986
- 6) BREITMAIER, E. & W. VOELTER (*Ed.*):  $^{13}\text{C}$  NMR Spectroscopy. pp. 74~75, Verlag Chemie, 1978
- 7) JANSEN, R.; G. REINFENSTAHL, K. GERTH, H. REICHENBACH & G. HOEFLE: Myxalamide A, B, C und D, Gruppe homologer Antibiotika aus *Myxococcus xanthus* Mx  $\times$  12 (Myxobacteriales). *Liebigs Ann. Chem.* 1983: 1081~1095, 1983